



Li StarFish S.r.l.
Via Cavour, 35 - 20063 Cernusco S/N (MI), Italy
Tel. +39-02-92150794 - Fax. +39-02-92157285
info@listarfish.it - www.listarfish.it

EDI™ Active GLP-1 (7-36) Specific ELISA Kit

Enzyme Linked ImmunoSorbent Assay (ELISA) for the measurement of active Glucagon-like peptide-1 (7-36) Level in Test Samples

Catalog Number: EPI-KT 871 Store at 2 – 8°C Upon Receipt

For Research Use Only.

INTENDED USE

This high sensitive ELISA (enzyme-linked immunosorbent assay) kit is produced for the exclusively quantitative determination of glucagon-like peptide-1 (7-36) [GLP-1 (7-36)] level in serum and EDTA-plasma sample among human, rat, mouse, goat, etc.

ASSAY PRINCIPLE

This ELISA is designed, developed and produced for the quantitative measurement of bioactive GLP-1 (7-36) in plasma sample. The assay utilizes the two-site “sandwich” technique with two selected GLP-1 (7-36) specific antibodies.

Assay standards, controls and test samples are directly added to wells of a microplate that is coated with streptavidin. Subsequently, a mixture of biotinylated GLP-1 (7-36) specific antibody and a horseradish peroxidase (HRP) conjugated GLP-1 (7-36) specific antibody is added to each well. After the first incubation period, a “sandwich” immunocomplex of “Streptavidin – Biotin-Antibody – GLP-1(7-36) – HRP conjugated antibody” is formed and attached to the wall of the plate. The unbound HRP conjugated antibody is removed in a subsequent washing step. For the detection of this immunocomplex, each well is then incubated with a substrate solution in a timed reaction and then measured in a spectrophotometric microplate reader. The enzymatic activity of the immunocomplex bound to GLP-1 (7-36) on the wall of the microtiter well is directly proportional to the amount of GLP-1 (7-36) in the sample.

REAGENTS: Preparation and Storage

This test kit must be stored at 2 – 8°C upon receipt. For the expiration date of the kit refer to the label on the kit box. All components are stable until this expiration date.

Prior to use allow all reagents to come to room temperature.

Reagents from different kit lot numbers should not be combined or interchanged.

1. Streptavidin Coated Microplate (Cat. No. 10040)

One well-breakable microplate with 12 x eight strips (96 wells total) coated with streptavidin. The plate is framed and sealed in a foil zipper bag with a desiccant. This reagent should be stored at 2 – 8°C and is stable until the expiration date on the kit box.

2. GLP-1 Tracer Antibody (Cat. No. 30229)

One vial containing 0.6 mL HRP labeled Anti-GLP-1 specific antibody in a stabilized protein matrix. This reagent must be mixed with GLP-1 (7-36) Capture Antibody and the tracer antibody diluent before use (for details see Assay Procedure). This reagent should be stored at 2 – 8°C and is stable until the expiration date on the kit box.

3. GLP-1 (7-36) Capture Antibody (Cat. No. 30230)

One vial containing 0.6 mL of biotinylated GLP-1 (7-36) specific antibody. It should be used only after mixed with GLP-1 Tracer Antibody and the tracer antibody diluent according to the assay procedures. This reagent should be stored at 2 – 8°C and is stable until the expiration date on the kit box.

4. ELISA Wash Concentrate (Cat. No. 10010)

One bottle contains 20 mL of 30 fold concentrate. Before use the contents must be diluted with 580 mL of distilled water and mixed well. Upon dilution this yields a working wash solution containing a surfactant in phosphate buffered saline with a non-azide and non-mercury based preservative. The diluted wash buffer should be stored at room temperature and is stable until the expiration date on the kit box.

5. ELISA HRP Substrate (Cat. No. 10020)

One bottle contains 22 mL of tetramethylbenzidine (TMB) with stabilized hydrogen peroxide. This reagent should be stored at 2 – 8°C and is stable until the expiration date on the kit box.

6. ELISA Stop Solution (Cat. No. 30357)

One bottle contains 12 mL of sulfuric acid. This reagent should be stored at 2 – 8°C or room temperature and is stable until the expiration date on the kit box.

7. GLP-1 Standards (Cat. No. 30261 – 30265)

Five vials containing different levels of lyophilized GLP-1 (7-36) in a liquid protein matrix with a non-azide, non-mercury based preservative. Refer to vial for exact concentration for each standard. These reagents should be stored at 2 – 8°C and are stable until the expiration date on the kit box.

8. GLP-1 Controls (Cat. No. 30266 – 30267)

Two vials containing different levels of lyophilized GLP-1 (7-36) in a liquid protein matrix with a non-azide, non-mercury based preservative. Refer to vials for exact concentration range for each control. Both controls should be stored at 2 – 8°C and are stable until the expiration date on the kit box.

9. Tracer Antibody Diluent (Cat. No. 30017)

One vial containing 12 mL ready to use buffer. It should be used only for tracer antibody dilution according to the assay procedures. This reagent should be stored at 2 – 8°C and is stable until the expiration date on the kit box.

SAFETY PRECAUTIONS

The reagents must be used in a professional laboratory environment and are for research use only. Source material (e.g. highly purified bovine serum albumin) of bovine serum was derived in the contiguous 48 United States. It was obtained only from donor healthy animals maintained under veterinary supervision and found free of contagious diseases. Wear gloves while performing this assay and

handle these reagents as if they are potential infectious. Avoid contact with reagents containing TMB, hydrogen peroxide, or sulfuric acid. TMB may cause irritation to skin and mucous membranes and cause an allergic skin reaction. TMB is a suspected carcinogen. Sulfuric acid may cause severe irritation on contact with skin. Do not get in eyes, on skin, or on clothing. Do not ingest or inhale fumes. On contact, flush with copious amounts of water for at least 15 minutes. Use Good Laboratory Practices.

MATERIALS REQUIRED BUT NOT PROVIDED

1. Precision single channel pipettes capable of delivering 25 µL, 50 µL, 100 µL, and 1000 µL etc.
2. Repeating dispenser suitable for delivering 100 µL.
3. Disposable pipette tips suitable for above volume dispensing.
4. Disposable 12 x 75 mm or 13 x 100 glass/plastic tubes.
5. Disposable plastic 100 mL and 1000 mL bottle with caps.
6. Aluminum foil.
7. Deionized or distilled water.
8. Plastic microtiter well cover or polyethylene film.
9. ELISA plate shaker
10. ELISA multichannel wash bottle or automatic (semi-automatic) washing system.
11. Spectrophotometric microplate reader capable of reading absorbance at 450 nm.
12. DPP-4 Inhibitor

SPECIMEN COLLECTION

Although both serum and EDTA-plasma samples can be used for measuring Active GLP-1 (7-36) using this assay, it is recommend to use EDTA-plasma, because the plasma sample showed a better stability than serum sample.

- (1) Only 200 µL of EDTA-plasma is required for bioactive GLP-1 (7-36) measurement in duplicate. More sample volume is needed if a sample extraction procedure is required.
- (2) No special preparation of individual is necessary prior to specimen collection. However, fasting sample and non-fasting/glucose induced sample may present great significance for bioactive GLP-1 (7-36) level.
- (3) Whole blood should be collected into a lavender top Vacutainer® EDTA-plasma tube. Invert tube to mix well and place the tube on ice. Centrifuge the tube within an hour at 1000 g for 10 minutes in a refrigerated centrifuge.
- (4) EDTA-plasma samples should be stored at 2 – 8°C if they will be tested within 3 hours of collection. For longer storage, it is recommended to store the plasma sample at -70°C. Avoid more than three times repeated freezing and thawing cycles. Aliquot samples before freezing if necessary.
- (5) It is very important to add appropriate amount of DPP-4 inhibitor to the collected specimen right after the separation of plasma from the blood cells. Refer to DPP-4 manufacturer's instruction.
- (6) BD™ P700 Blood Collection and Preservation System (contains a DPP-4 protease inhibitor cocktail) is recommended. However, each institute should verify the collection system for their specific research and study.

ASSAY PROCEDURE

1. Reagent Preparation

- (1) Prior to use allow all reagents to come to room temperature. Regents from different kit lot numbers should not be combined or interchanged.
- (2) ELISA Wash Concentrate must be diluted to working solution prior use. Please see REAGENTS section for details.
- (3) Reconstitute all standards and controls by adding **1.0 mL** of demineralized water to each vial. Allow the standards

and controls to sit undisturbed for 10 minutes, and then mix well by gentle vortexing. These reconstituted standards and controls must be stored at -20°C or below. Do not exceed 3 freeze-thaw cycles.

2. Test Sample Preparation

Although EDTA-plasma or serum samples can be directly measured for the Active GLP-1 (7-36) concentration, some studies using this assay by pharmaceutical companies indicated that sample pre-treatment with a column extraction is necessary to obtain an accurate and clinical meaningful test results.

Epitope Diagnostics provides a validated and user friendly column extraction procedures and reagents packed as a GLP-1 sample extraction kit (Catalog No. KT-910). This kit provides all the reagents ready to use. This extraction procedure does not require any special equipment, such as vacuum centrifuge, etc.

3. Assay Procedure

- (1) Place a sufficient number of streptavidin coated microwell strips/wells in a holder to run GLP-1 (7-36) standards, controls and unknown samples in duplicate.
- (2) Test Configuration

ROW	STRIP 1	STRIP 2	STRIP 3
A	STD 1	STD 5	SAMPLE 2
B	STD 1	STD 5	SAMPLE 2
C	STD 2	C 1	SAMPLE 3
D	STD 2	C 1	SAMPLE 3
E	STD 3	C 2	SAMPLE 4
F	STD 3	C 2	SAMPLE 4
G	STD 4	SAMPLE 1	
H	STD 4	SAMPLE 1	

- (3) Prepare GLP-1 (7-36) Antibody Mixture: mixing GLP-1 Tracer Antibody and Capture Antibody by 1:21 fold dilution of the Tracer Antibody (30229) and by 1:21 fold dilution of the biotinylated Capture Antibody (30230) with the Tracer antibody Diluent. For each strip, it is required to mix 1 mL of the Tracer Antibody Diluent (30017) with 50 µL the Capture Antibody and 50 µL of the Tracer Antibody in a clean test tube.
- (4) Add **100 µL** of standards, controls and test samples into the designated microwell.
- (5) Add **100 µL** of GLP-1 (7-36) Antibody Mixture to each well
- (6) Cover the plate with one plate sealer and incubate plate at 2-8°C, static for **20 - 24 hours**.
- (7) Remove plate sealer. Aspirate the contents of each well. Wash each well 5 times by dispensing 350 µL of working wash solution into each well and then completely aspirating the contents. Alternatively, an automated microplate washer can be used.
- (8) Add **200 µL** of ELISA HRP Substrate into each of the wells.
- (9) Cover the plate with one plate sealer and also with aluminum foil to avoid exposure to light.
- (10) Incubate plate at room temperature, static for **20 min**.
- (11) Remove the aluminum foil and plate sealer. Add **50 µL** of ELISA Stop Solution into each of the wells. Mix gently.
- (12) Read the absorbance at **450nm/620 nm** within 10 minutes in a microplate reader

NOTE: to reduce the background, one can set the instrument to dual wavelength measurement at 450 nm with background wavelength correction set at 595 nm or 620 nm or 630 nm.

PROCEDURAL NOTES

1. It is recommended that all standards, controls and unknown samples be assayed in duplicate. The average absorbance reading of each duplicate should be used for data reduction and the calculation of results.
2. For samples with higher than level 5 standard, it is recommended to measure diluted the specimen with an appropriate GLP-1 free human serum matrix for a more accurate report.
3. Keep light sensitive reagents in the original amber bottles.
4. Store any unused streptavidin coated strips in the foil zipper bag with desiccant to protect from moisture.
5. Careful technique and use of properly calibrated pipetting devices are necessary to ensure reproducibility of the test.
6. Incubation times or temperatures other than those stated in this insert may affect the results.
7. Avoid air bubbles in the microwell as this could result in lower binding efficiency and higher CV% of duplicate reading.
8. All reagents should be mixed gently and thoroughly prior use. Avoid foaming.

INTERPRETATION OF RESULTS

1. Calculate the average absorbance for each pair of duplicate test results.
2. Subtract the average absorbance of the STD 1 (0 ng/mL) from the average absorbance of all other readings to obtain corrected absorbance.
3. The standard curve is generated by the corrected absorbances of all standard levels on the ordinate against the standard concentration on the abscissa using point-to-point or log-log paper. Appropriate computer assisted data reduction programs may also be used for the calculation of results. We recommend using **Point-to-Point** or **Quadratic** curve fit.

The GLP-1 (7-36) concentrations for the controls and test samples are read directly from the standard curve using their respective corrected absorbance. If log-log graphic paper or computer assisted data reduction program utilizing logarithmic transformation are used, sample having corrected absorbance between the 2nd standard and the next highest standard should be calculated by the formula:

$$\text{Value of unknown} = \frac{\text{Corrected absorbance (unknown)}}{\text{Corrected Absorbance (2}^{\text{nd}} \text{ STD)}} \times \text{Value of the 2}^{\text{nd}} \text{ STD}$$

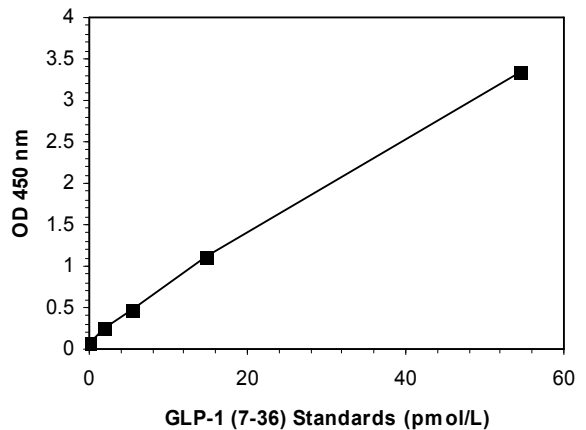
EXAMPLE DATA AND STANDARD CURVE

A typical absorbance data and the resulting standard curve from this GLP-1 ELISA are represented. **This curve should not be used in lieu of standard curve run with each assay.**

Well I.D.	OD 450 nm Absorbance			Results pmol/L
	Readings	Average	Corrected	
0 pmol/mL	0.069 0.082	0.076	0.000	
1.82 pmol/L	0.240 0.271	0.255	0.179	
5.46 pmol/L	0.464 0.491	0.477	0.401	
14.74 pmol/L	1.105 1.101	1.103	1.027	
54.58 pmol/L	3.353 3.341	3.347	3.271	

Control I	0.387 0.367	0.377	0.301	4.1
Control II	0.996 1.005	1.000	0.924	13.2

Active GLP-1 (7-36) Specific ELISA



EXPECTED VALUES

Each laboratory should establish its own normal range by using samples collected from normal healthy people. Please be note that the normal range may be variable by using fasting samples vs. non-fasting samples.

GLP-1(7-36) pg/ml = GLP-1 (7-36) pmol/l x 3.298

LIMITATION OF THE PROCEDURE

1. Since there is no Gold Standard concentration or international standard available for GLP-1 (7-36) measurement, the values of assay standards were established using a highly purified GLP-1 (7-36) peptide and validated by Epitope Diagnostics. Results obtained with different assay methods or kits cannot be used interchangeably.
2. For unknown sample value read directly from the assay that is greater than assay standard level-5, it is recommend measuring a diluted sample for more accurate measurement.
3. Bacterial or fungal contamination of serum specimens or reagents, or cross contamination between reagents may cause erroneous results.
4. Water deionized with polyester resins may inactivate the horseradish peroxidase enzyme.

QUALITY CONTROL

To assure the validity of the results each assay should include adequate controls with known GLP-1 (7-36) levels.

PERFORMANCE CHARACTERISTICS

Sensitivity

The sensitivity of this High Sensitive GLP-1 (7-36) ELISA as determined by the 95% confidence limit on 12 duplicate determination of zero standard is about 0.3 pmol/L. By linear dilution of assay standard level 2 (1.82 pmol/L) to 0.91 pmol/L and 0.46 pmol/L, the assay can clearly detect both of them. Therefore, a realistic detection limit of 0.5 pmol/L is easily achieved by this assay.

Specificity

This Bioactive GLP-1 (7-36) assay is specific measure GLP-1 (7-36). It is expected that this assay does not detect following peptides.

GLP-1 (7-36)	100%
GLP-1 (9-36)	< 0.1%
GLP-1 (9-37)	< 0.1%
GLP-1 (7-37)	< 0.1%
GLP-1 (1-36)	< 0.1%
GLP-2	< 0.1%
Glucagon	< 0.1%

Linearity

Two samples were diluted with GLP-1 (7-36) zero human serum matrix. These diluted samples are measured in this assay and the linear recovery is calculated.

Sample Dilution format	GLP-1 (7-36) zero human serum	Measured value (pmol/L)	Expect value (pmol/L)	Linear recovery (%)
Sample 1				
50 µl	250 µl	0.64	0.70	91.3
100 µl	200 µl	1.06	1.39	76.1
150 µl	150 µl	2.21	2.12	104.3
200 µl	100 µl	2.88	2.82	102.2
250 µl	50 µl	4.18	3.55	117.9
Sample 2				
50 µl	250 µl	2.70	2.79	96.7
100 µl	200 µl	5.09	5.55	91.8
150 µl	150 µl	7.49	8.34	89.8
200 µl	100 µl	11.40	11.13	102.5
250 µl	50 µl	13.37	13.89	96.3

Spike Recovery

Patient samples were spiked each other in the sample volume (200 µl + 200 µl) and measured with this assay. The spike recovery are calculated.

Samples (pmol/L)	Samples (pmol/L)	Measured value (pmol/L)	Expect value (pmol/L)	Recovery (%)
32.45	7.49	18.56	19.98	92.9
6.67	18.62	11.25	12.65	89.0
2.40	7.61	4.12	5.00	82.4

WARRANTY

This product is warranted to perform as described in its labeling and literature when used in accordance with all instructions. Epitope Diagnostics, Inc. DISCLAIMS ANY IMPLIED WARRANTY OF MERCHANTABILITY OR FITNESS FOR A PARTICULAR PURPOSE, and in no event shall Epitope Diagnostics, Inc. be liable for consequential damages. Replacement of the product or refund of the purchase price is the exclusive remedy for the purchaser. This warranty gives you specific legal rights and you may have other rights, which vary from state to state.

REFERENCES

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Short Assay Protocol:

- Add 100 µl/well of standards, control and patient sample
- Add 100 µl of Antibody Mixture
- Incubate 20 - 24 hour at 2-8°C, static
- Wash strips with diluted wash buffer
- Add 200 µl/well of TMB substrate
- Incubate 20 min at RT, static
- Add 50 µl stop solution
- Read strips at OD 450 nm/620 nm